

Received: July 17, 2023**Published:** Aug 18, 2023

Evalution of Antibacterial Activity of Methanol leaf and stem Bark Extracts of Newboulia laevis, Parkia biglobosa And Conventional Available Antibiotics on Methicillin Resistant *Staphylococcus aureus*

Ikemesit Peter^{4*}, Veronica Nwode¹, Kayode Akindele¹, Nnyeneime Bassey², Evangeline Chinyere Udenweze³, Lynda Ugwu¹, Ijeoma Onyinye Okolo², Chinene Emelda Nwankwo³, Farotimi Chinene Asumpta¹, Ikechukwu Obya¹, Augustina Nwatu¹, Henrietta Onu¹, Ifeanyichukwu Romanus Iroha⁵

¹Department of Microbiology, Faculty of Biological Science, Alex Ekwueme Federal University Ndifur-Alike, P.M.B. 1010, Ikwo, Ebonyi State, Nigeria.

²Department of Applied Science, Faculty of Pure and Applied Science, Federal College of Dental Technology and Therapy, Trans Ekulu, P.M.B. 01473, Enugu, Nigeria.

³Department of Applied Microbiology and Brewing, Faculty of Bioscience, Nnamdi Azikiwe University, Awka, Anambra, P. M. B. 5025, Nigeria.

⁴Department of Public Health, Faculty of Health Technology and Engineering, Federal College of Dental Technology and Therapy, Trans-Ekulu, P.M.B. 01473, Enugu, Nigeria.

⁵Department of Applied Microbiology, Faculty of Science, Ebonyi State University, Abakaliki, P.M.B. 53, Nigeria.

Abstract

Background: Methicillin-resistant *Staphylococcus aureus* (MRSA) strains continue to be a leading cause of infections with high morbidity and mortality in wound patients, but little is known about the prevalence, characterization, and evaluation of antibacterial activity of methanol leaf and stem bark extracts of Newbouldia laevis, Parkia biglobosa and conventional available antibiotics on MRSA from various wound sources in Abakaliki. This study aimed at investigating the prevalence of MRSA and its associated risk factors amongst wound patients in a Tertiary Hospital. **Methods:** A total of 200 samples from post-surgical wounds, burn wounds, decubitus ulcer and diabetic foot ulcers patient were collected from March 2021 to February 2022 using a sterile swab sticks on the affected site. The collected specimens were immediately subjected to bacteriological analysis. MRSA were screened for Antibiotic Susceptibility Testing (AST) and results were analyzed using the Clinical Laboratory Standard Institute (CLSI) zone diameter breakpoints. Antibacterial activity of methanol leaf and stem-bark extract of Newbouldia laevis and P. biglobosa against the MRSA isolates were determined using agar well diffusion Method. **Results:** the prevalence of MRSA accounted 55.5 %. Antibiogram result shows that the MRSA isolate were 87.8% and 100% susceptible to imipenem and amikacin but 78.9% and 100% resistant to erythromycin, penicillin, lincomycin, vancomycin, trimethoprim-sulfamethoxazole, tetracycline respectively. At 25mg/ml, 12.5mg/ml and 6.25mg/ml, Parkia biglobosa showed high inhibitory activity against MRSA at the range of 9mm – 19mm compared to Newbodia leavis having inhibitory activity against MRSA at the range of 0 – 15mm at the same concentrations. **Conclusion:** Our study report the prevalence of MRSA with multi-resistant in wound samples and also advocate for judicious use of imipenem and amikacin as drug of choice against the test isolate. This findings validate the use of methanol leaves and stem-bark extract of P. biglobosa as alternative medicine for the treatment of MRSA-associated wound infection and recommends further complementary studies in ethno pharmacological use.

Keywords: Methicillin-resistant *Staphylococcus aureus*, Parkia biglobosa, Newbodia leavis, wound.

Introduction

Methicillin Resistant *Staphylococcus aureus* (MRSA) are strain encoding MecA gene which mediate resistance to penicillin-derived methicillin and other beta-lactam antibiotic [1]. The presence of an altered penicillin binding protein (PBP2a) encoded by MecA strain has been recognized as factor essential for methicillin resistant [2]. This house-keeper gene is found as an integral portion of the *Staphylococcal* cassette chromosome mec (SCC mec) [2, 3]. Methicillin Resistant *Staphylococcus aureus* are considered as priority pathogen in community and hospital acquired infection of which their potential risk are also matters of concern [2].

MRSA has a somewhat high global burden in immunocompromised patients particularly wound patients causing significant morbidity and mortality [4]. In human, MRSA remains one of the medically important bacteria that colonizes bone and wound infections. The expression of virulence by MRSA in osteomyelitis and postoperative wound infections orthopedic medicine [6] are the leading cause of delayed bone/wound healing and non-union which may necessitate the amputation of such bones [2, 6]. Prior study has reported high risk of MRSA infection in fracture and post-surgical wound patients [6]. MRSA are often multi-resistant eroding the effective of most conventional antibiotic with limited treatment option. Another key element that contributes to treatment failure in wound patients is the creation of various virulence factors. For example, MRSA strains that produce Panton-Valentine Leukocidin (PVL) have higher lethality than PVL-negative bacteria [6] as observed in soft tissue infection, surgical site infection (SSI) and burn wound [7].

Despite the fact that MRSA is a substantial health issue, there is a lack

***Corresponding Author:** *Ikemesit Peter, Department of Public Health, Faculty of Health Technology and Engineering, Federal College of Dental Technology and Therapy, Trans-Ekulu, P.M.B. 01473, Enugu, Nigeria.

Citation: *Ikemesit Peter. Evalution of Antibacterial Activity of Methanol leaf and stem Bark Extracts of Newbouldia laevis, Parkia biglobosa And Conventional Available Antibiotics on Methicillin Resistant *Staphylococcus aureus*. Jourl of Clin Stud and Med Imag, Cas Rep. 2023; 1(4): 1018.

of research on the frequency of MRSA in wounds in Nigeria. In most hospital department/ward with wound patients, Ampicillin, ampiclo, gentamicin, and ciprofloxacin are the most widely prescribed antibiotics. The majority of recently utilized medications in most hospital are not novel in therapeutic use, but rather refinements of existing classes of antibacterial treatments that can only provide temporary relief to wound patients.

It is, nonetheless, difficult to imagine a world without antibiotics. It is, nonetheless, difficult to imagine a world without antibiotics. However, this is rapidly approaching because diseases and disease-causing agents that were formerly thought to have been conquered and beaten by antibacterial medicines are emerging in a new pattern of resistance to therapeutic drugs.

Researchers are currently looking for compounds from plants that can operate as cutting-edge antimicrobials, with a broader spectrum of activity against bacteria of both Gram-positive and Gram-negative origin and no major side effects [8, 9]. The ability of plants to produce an endless number of secondary metabolites that serve as the foundation for plant-derived antibacterial agents.

Smooth Newbouldia or boundary tree is the popular name for Newbouldia laevis (Bignoniaceae) [10]. It can reach a height of 7 - 15 meters, but it is more commonly a shrub of 2-3 meters with multiple stems producing clumps of gnarled branches [10]. Most tribes in Nigeria refer to it as Eto Adanha in Ibibio-Efik, Akoko in Yoruba "Ogiris" in Igbo, "Ikhimi" in Edo, and "Aduruku" in Hausa [10, 11]. Newbouldia laevis is commonly used in folk medicine in South Eastern and Midwestern Nigeria for the treatment of infected wounds and eye issues [10]. Scientific studies on the plant's phytochemical contents indicated the presence of phenylpropanoids and alkaloids in the root [12] tannins and flavonoids in the leaves [13]. Also, Parkia biglobosa (African locust bean tree) is native to Africa [14] and is commonly referred to as Orgili in the Igbo tribe of Nigeria. Parkia biglobosa is a plant that has demonstrated promising source of phytomedicinal compound [15], as well as other ethnobotanical and folkloric applications. The World Health Organization (WHO) has stated that medicinal plants may be the finest source of a wide range of medications [16]. As a result, there has been a global renaissance in the use of herbal remedies in illness management across all continents, with most developing countries now incorporating phytomedicine into their health care systems. Diseases are treated in Nigeria's health-care system utilizing medicinal plants alongside with contemporary medicine. As the looming threat of clinical antibacterial tolerance or resistance persists, susceptibility tests remain essential for the screening and selection of medicinal and conventional antibacterial agents. As a result, this study looks into the screening of available antibiotic and antibacterial ability of Parkia biglobosa and Newbouldia laevis leaf and stem bark as a panacea for the treatment of MRSA wound infection.

Methods

Inclusion and Exclusion Criteria

The study comprised consenting patients with current post-surgical wound, burn wound, decubitus ulcer, and diabetic foot ulcers occurring within 30 days or within one year if an orthopedic implant is in place. Patients with community-acquired pyogenic infections such as furuncle, abscess, and carbuncle; patients with episiotomy infection; and patients with open fractures were excluded from the study.

Demographic Data and Sample collection

Demographic and clinical data were acquired from consenting patients utilizing a standardized validated questionnaire and patient files. These included age, gender, and previous antibiotic intake, beginning of the lesion/length of hospital stay, type, and associated medical condition. A total of 200 samples (Fifty [50] each) were col-

lected within the period of 12 months (March 2021 to February 2022) from post-surgical wound, burn wound, decubitus ulcer and diabetic foot ulcers patients were collected by rotating a sterile swabs on the affected site. The collected specimens were taken within one hour of collection via moist Amie's transport medium to the microbiology research unit of the Ebonyi State University Abakaliki, Nigeria for bacteriological investigation.

Isolation and identification of test organisms

The wound swab specimens were suspended in a sterilized nutrition broth (Oxoid, UK) and incubated for 24 hrs at 37oC. The turbid bacterial growth was plated onto MP1571 Denim Blue Chromogenic MRSA Screening Agar (Oxoid, UK), and incubated for 24 hrs at 37oC. Bacterial colonies on MP1571 Denim Blue Chromogenic MRSA Screening Agar (Oxoid, UK) with atypical MRSA features of denim blue colonies were aseptically purified by subculturing onto nutrient agar (Oxoid, UK) and incubated at 24 hrs at 37oC. The pure culture strain were subsequently identified via and PBP2 latex agglutination tests (DR0900A) and Vitex 2 compact 60 next-generation automated system (BIOMERIEUX, U.S.A).

Antibiotic Susceptibility Testing

According to the Clinical and Laboratory Standards Institute (CLSI) criteria (CLSI, 2019), a standardized Kirby-Bauer disk diffusion method with the Mueller-Hinton agar (MHA) (Oxoid, Basingstoke, Hampshire, UK) plate technique was performed. A suspension of bacterial cells (i.e., equivalent of 1x10⁶ colony forming unit per milliliter (cfu/ml) was swabbed onto each Petri dish containing solidified Mueller-Hinton agar and adjusted to 0.5 MacFarland turbidity standard. The inoculated organisms was allowed for 15 minutes for pre-diffuse of bacteria suspension. Antibiotic discs that are commercially available and comprising the following antibiotics: Vancomycin (30 µg), Amikacin (10µg), Ceftazidime (30 µg), Cefotaxime (30 µg), Erythromycin (5µg), lincomycin (15µg), Imipenem (30 µg), Trimethoprim-Sulfamethoxazole (30 µg), Penicillin (10 µg), Tetracycline (15µg) (Oxoid) were was aseptically applied with sterile forceps onto the surfaces of the solidified Mueller-Hinton plates and gently pressed to achieve uniform contact. The plates was incubated for 24 hrs at 37° C. The Inhibition zones were interpreted in millimeter (mm) according to CLSI guideline for antibiotics sensitivity [2, 16].

Collection and Authentication of Parkia biglobosa and Newbouldia laevis

Parkia biglobosa and Newbouldia laevis leaf and stem bark were collected at Ndume-Alike community in Ikwo L. G. A in Ebonyi State. The plants materials were examined and validated with voucher no: HP/344/67R by Dr. C. D Udechukwu, a taxonomist in the Department of Biology at Alex Ekwueme Federal University Ndume-Alike in Ikwo L. G. A in Ebonyi State, Nigeria.

3.1.25 Extraction of Stem-bark and Leaf of Parkia biglobosa and Newbouldia laevis

This was performed following a modification in methods of Osuntokun et al. [17] as stated below.

Modification: The work of Osuntokun et al. [17] air-dried the plant material for 10days and the plant materials were not cut into pieces before maceration. The filtrate was dried using a rotary evaporator. Parkia biglobosa and Newbouldia laevis leaf and stem bark were sun-dried for 5 days after being washed with sterile water. The plant material were cut into pieces and then pulverize with a sterile mortar and pestle to reduce into a powdered particles [14, 17]. Prior to extraction, exactly 200 grams of Parkia biglobosa and Newbouldia laevis each leaf and stem bark was soaked in 400 ml volume of methanol and allowed to stand for overnight at room temperature. It was then filtered with muslin cloth [8]. The filtrate was evaporated to dryness at <40oC using rotary evaporator and stored in the desiccator until needed.

Preparation of Different Concentration of the Extracts

Exactly, one gram (1g) of each plant part was reconstituted separately in 10ml of 70% Dimethylsulphoxide as diluent to obtain a concentration of 100mg/ml. Exactly, 10-5 serial dilution was carried out from the stock solution to obtain extract concentration of 50 mg/ml, 25 mg/ml, 12.5 mg/ml and, 6.25 mg/ml.

Antibacterial Activity of Methanol stem-bark and leaf extract of *Parkia biglobosa* and *Newbouldia laevis*

The antibacterial activity of plant extracts was determined using the Agar well diffusion method as described by earlier researchers [8, 18]. A suspension of bacterial cells (i.e., equivalent of 1x10⁶ colony forming unit per milliliter (cfu/ml) was seeded onto each Petri dish containing solidified Mueller-Hinton agar and adjusted to 0.5 MacFarland turbidity standard. This was allowed to stand for 15 minutes to enable the inoculated organisms to pre-diffuse. The wells were then aseptically bored on seeded agar plates with an 11 mm sterile Cork borer. The wells were filled with 1ml of each extract concentration. The inoculated agar plates were incubated for 24 hours at 37OC. After incubation, clear inhibition zones were measured using a metric rule and interpreted in millimeters (mm).

Result

Prevalence of MRSA from wound samples

MRSA from isolated *Staphylococcus aureus* recovered from post-surgical wound, burn wound, diabetic foot ulcers and decubitus ulcer samples showed prevalence rate of 87.2, 78.5, 51.7, and 62.9 respectively. The total prevalence rate was 55.5 % as shown in Table 1.

Demographic and Relevant Clinical information of wound patients with MRSA

MRSA among wound patients were predominant amongst 35-49years recording 21.5%, followed by 50 – 69 years recording 18.5% and 20 – 34 years having 15.5%. Gender distribution of MRSA was observed in Female than male with MRSA occurrence rate of 33.5% and 22.0% respectively. Duration of hospitalization within 2months displayed high prevalence of MRSA 18.5%. Duration of 4 months and above showed 10.5%, Greater than 1 week 10.0, 7 days showed 7.5%, Duration of 3 months showed 5.0% and the least prevalence of MRSA was duration of 1 month as shown in Table 2.

Antibiotic susceptibility profile of MRSA

The result of antibiotic susceptibility profile of MRSA from post-surgical wound swabs in Table 2 showed 100%, 87.8%, 51.2% susceptibility to Imipenem, Amikacin and Cefotaxime respectively. The isolates also showed highest (100%) resistance to Penicillin and tetracycline, followed by 97.5%, 951% and 87.8% resistance to Lincomycin, trimethoprime-sulfamethoxazole and Vancomycin.

MRSA isolated from burn wound patient were 100% resistant to penicillin, tetracycline, trimethoprim-Sulfamethoxazole, Vancomycin while 78.9%, 69.7% and 57.6% resistant was observed against erythromycin, lincomycin and ceftazidime respectively. The MRSA isolated was 100% susceptible to Amikacin, imipenem and Cefotaxime followed by 42.2%, 30.3% and 21.2% recorded against Ceftazidime, lincomycin and Erythromycin respectively. While Amikacin, cefotaxime and imipenem showed 100% susceptible to the MRSA isolates from burn wound patients, ceftazidine, erythromycin and lincomycin recorded 42.4, 21.2 and 30.3 susceptible respectively against MRSA isolate from burn wound as shown the Table 2.

MRSA recovered from decubitus ulcer patient were susceptible to aminoglycoside (amikacin 90.9%), cephalosporin (cefotaxime 100%) and Cabapenem (imipenem 100%) but less susceptible against erythromycin and ceftazidime recording 13.6% and 27.3% respectively. MRSA isolated from Decubitus ulcer Patient exhibit high level of resistance recording 100% to Trimethoprim-Sulfamethoxazole, Vancomycin, Tetracycline, Penicillin and Lincomycin while 86.4%, 72.7%

and 9.1% resistance rate was recorded against Erythromycin, Ceftazidime and Amikacin respectively as presented in Table 3. Also, MRSA recovered from Diabetic foot ulcers patient were 100% resistance to lincomycin, penicillin, tetracyclin, trimethoprim – sulfamethoxazole and vanconycin. Amikacin showed resistance of 9.1 against 90.9 susceptible. Then, cetrizidine and erythromycin recorded 72.7 and 86.4 resistance as presented in the Table 3.

Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from post-surgical wound

Methanol leaf extract of *Parkia biglobosa* had inhibition zone diameter within the range of 11-15mm at 100mg/ml concentration while stem-bark extract had inhibition zone diameters within the range of 19-26 mm at 100mg/ml concentration against MRSA isolates from post-surgical wound in Table 4.

Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from Burn wound

Methanol leaf extract of *Parkia biglobosa* showed no inhibitory effect against MRSA isolates from burn wound while stem-bark extract showed antibacterial activity at 100mg/ml concentration within the range of 15-25mm against the test isolates (Table 5). stem-bark extract demonstrated antibacterial activity within the range of 13-21mm and 9-17mm at 50mg/ml and 25mg/ml concentration against MRSA isolates from burn wound.

Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from Burn wound

MRSA isolates from Diabetic foot ulcers were less sensitive to methanol leaf extract of *Parkia biglobosa* at 12.5 mg/ml and 6.25 mg/ml concentration with no inhibitory effect while high inhibitory zone diameter of 24mm and 21mm was observed at 100mg/ml and 50 mg/ml concentration of stem-bark extract as presented in Table 6.

Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from Decubitus ulcer

Stem-bark extract of *Parkia biglobosa* was active at 6.25 mg/ml concentration recording 9-11mm against MRSA isolates from Decubitus ulcer. While the stem bark showed active resistance at concentration 100mg/ml recording 14-25mm and 9-21mm at the concentration 50mg/ml as shown in table 7.

Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Post-surgical wound

Methanol crude extract of *Newbouldia laevis* showed no inhibitory activity at 12.5 mg/ml and 6.25 mg/ml concentration against MRSA isolates from post-surgical wound .The leaf extract at 100mg/ml showed highest activity measuring 18mm but the stem bark recorded high activity measuring 20mm as shown in Table 8.

Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Burn wound

Leaf extract of *Newbouldia laevis* demonstrated high antibacterial activity with inhibitory zone diameter of 15mm and 12mm at 100mg/ml and 50 mg/ml concentration respectively while *Newbouldia laevis* stem-bark extract showed no inhibitory activity at 12.5 mg/ml and 6.25 mg/ml concentration against MRSA isolates from burn wound shown in Table 9.

Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Diabetic foot ulcers

Newbouldia laevis leaf and stem bark extract against MRSA isolates from diabetic foot ulcers revealed inhibitory zone diameter 20mm, 17mm and 15mm and 15mm at 100mg/ml and 50 mg/ml concentration respectively as shown in Table 10.

Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Decubitus ulcer

Methanol leaf extract of *Newbouldia laevis* demonstrated an inhibitory effect against MRSA isolates from decubitus ulcer while Stem-bark extract revealed high inhibitory zone diameter of 21mm and

17mm at 100mg/ml and 50 mg/ml concentration respectively (Table 11).

Discussion

MRSA is assessed by a series of resistant determinant according to different strains. As a result, the chromogenic agar was used to identify MRSA strain in wound with occurrence rate of 55.5%. In concordance with this result, high incidence of MRSA has been found in wound infection in Nigeria; Olowe et al. [19] reported 47.7% in Osogbo, south-western Nigeria [19], two studies reported 75% in Zaria [1, 20], Ikeh. [21] reported an MRSA prevalence of 81% from in-patients in Jos, Nigeria. Nwankwo and Nasiru [22] reported 62% prevalence rate from in-patients in kano Nigeria. In Enugu, Nigeria, MRSA strain accounted 86(28.4 %) and 78(25.7 %) from post-surgical wound and fracture wound respectively [2]. Taiwo et al. [23], earlier reported a prevalence of 70.6% in the city of Ilorin from in-patients.

However, low and higher values have been reported in wound samples from inter-country studies: 18.8% in Mwanza-Tanzania [24], 25% in Jinja-Uganda [25], 37.4% in Madinah kingdom of Saudi Arabia

[26], 25 (61%) in Iran [4], 75% from surgical wounds in Algeria [27], 72.0% in Eritrea [5], 80% in Peru [28] and in a setting in Colombia 90% [29]. Many published research have also documented a rise in the prevalence of MRSA, the majority of which was caused by wounds [30, 31]. Several reasons can explain the intra- and inter-country heterogeneity in MRSA prevalence. These include variances in study design, specimen type, laboratory techniques, and study duration, among other things [32]. Some studies, for example, depend completely on phenotypic processes to detect MRSA, whilst others rely on DNA-based approach (Multilocus Sequence Typing [MLST]), microarrays, whole genome sequencing, spa-typing, conventional PCR and, among others) to identify *mecA*. Apparently, investigations relying on DNA-based technique by PCR tends to reveal a lower incidence of MRSA [22, 31].

MRSA was more predominance in Post-surgical wound 20.5% than other wound sample in the study. This reiterate with the prevalence of MRSA 3.9%-35.6% in Post-surgical wound documented by other researchers [5, 33]. Our findings suggest that post-surgical patients may be prone to toxigenic equipment carriage and antibiotic-resistance clonal strains of *S. aureus*. More research into the pathogenicity potential and characteristics of MRSA populations from post-surgical wound patients could be beneficial. Given the discovery of numerous toxin genes, including the *tst* gene, in hospitalized post-operative patients [33]. Post-surgical wound patients in this study may likely be at risk of postsurgical toxic shock syndrome which result in delay wound healing and prolong hospitalization.

In this study, male patients harbor MRSA than female with MRSA occurrence rate of 33.5% and 22.0% respectively and is consistent with earlier research: male 55.9% vs female 44.1% [5], male 71% vs female 25% [26]. This is also consistent with previous studies from other parts of the world. In one such study from Madinah it was reported that MRSA strains were three times more prevalence from male patients in all studied cases [34, 35]. A study from Makkah found a similar pattern of males having higher MRSA isolation rates [36]. This trend could be linked to a variety of factors, including males being more susceptible to bacterial sepsis; being less compliant and so inclined to greater infection rates [37]. According to research, the female hormone (estrogen) has a negative effect on the development of virulence factors in pathogens [38]. Furthermore, the patients' personal hygiene habits, socioeconomic status, occupation and lifestyle all play essential roles in determining gender-infection correlations, which must be investigated further. It is generally known that women make up a larger proportion of the workforce in Abakaliki, so it is not surprising that samples taken from female patients were more numerous, resulting in a low MRSA isolation rate.

Age group assessment of MRSA among wound patients were more predominant amongst 35-49years recording 21.5%. Another study has reported high rate of MRSA with 22(37.3%) in 19–40year age group [5]. Low prevalence in this study was observed amongst age

Table 1: Prevalence of MRSA from wound samples.

Clinical Sample	No. of sample	MRSA positive (%)
Post-surgical wound	67	41(20.5)
Burn wound	51	33(16.5)
Diabetic foot ulcers	35	15(17.5)
Decubitus ulcer	47	22(11.0)
Total	200	111(55.5)

Table 2: Demographic and Relevant Clinical information of wound patients with MRSA.

Characteristic	No. sampled	MRSA (%)
Age group	20-34years	68 31(15.5)
	35-49years	84 43(21.5)
	50-69 years	48 37(18.5)
Gender	Male	96 67(33.5)
	Female	104 44(22.0)
Duration of hospitalization	7 days	26 15(7.5)
	>1week	39 20(10.0)
	1 month	27 8(4.0)
	2months	53 37(18.5)
Medical Condition	3months	24 10(5.0)
	4months & above	31 21(10.5)
	Post-surgical wound	67 41(87.2)
Medical Condition	Burn wound	51 33(78.5)
	Decubitus ulcer	47 22(62.9)
	Diabetic foot ulcers	35 15(51.7)
Previous Anti-biotic intake	Ampicillin	56 29(14.5)
	Ampiclox	67 47(23.5)
	Ciprofloxacin	43 15(7.5)
	Gentamicin	34 20(10.0)

Table 3: Antibiotic susceptibility profile of MRSA.

Antibiotic	Disc po-tency (µg)	Post-surgical wound n=41		Burn wound n=33		Decubitus ulcer n=22		Diabetic foot ulcers n=15	
		R (%)	S (%)	R (%)	S (%)	R (%)	S (%)	R (%)	S (%)
Amikacin	20	5(12.2)	36(87.8)	0(0.0)	33(100)	2(9.1)	20(90.9)	0(0.0)	15(100)
Cefotaxime	30	20(48.9)	21(51.2)	0(0.0)	33(100)	0(0.0)	22(100)	7(46.7)	8(53.3)
Ceftazidime	30	24(58.5)	17(41.5)	19(57.6)	14(42.4)	16(72.7)	6(27.3)	12(80)	3(20)
Erythromycin	15	21(51.2)	20(48.9)	26(78.9)	7(21.2)	19(86.4)	3(13.6)	15(100)	0(0.0)
Imipenem	30	0(0.0)	41(100)	0(0.0)	33(100)	0(0.0)	22(100)	0(0.0)	15(100)
Lincomycin	15	40(97.5)	1(2.4)	23(69.7)	10(30.3)	22(100)	0(0.0)	15(100)	0(0.0)
Penicillin	10	41(100)	0(0.0)	33(100)	0(0.0)	22(100)	0(0.0)	15(100)	0(0.0)
Tetracycline	30	41(100)	0(0.0)	33(100)	0(0.0)	22(100)	0(0.0)	10(66.7)	5(33.3)
Trimethoprim-Sulfa-methoxazole	30	39(95.1)	2(4.9)	33(100)	0(0.0)	22(100)	0(0.0)	15(100)	0(0.0)
Vancomycin	30	36(87.8)	5(12.2)	33(100)	0(0.0)	22(100)	0(0.0)	15(100)	0(0.0)

Key: R-Resistance , S- Susceptible, n-number of MRSA.

Table 4: Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from post-surgical wound.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 2	Leaf	11	10	NILL	8	7
MRSA 5		NILL	NILL	NILL	NILL	NILL
MRSA 7		NILL	NILL	NILL	NILL	NILL
MRSA 9		15	12	10	7	8
MRSA10		14	9	9	8	NILL
MRSA 11		14	12	10	9	9
MRSA 12		13	12	8	NILL	NILL
MRSA 14		NILL	NILL	NILL	NILL	NILL
MRSA 15		NILL	NILL	NILL	NILL	NILL
MRSA 2		21	20	19	NILL	NILL
MRSA 5		20	15	NILL	NILL	NILL
MRSA 7		21	17	12	NILL	NILL
MRSA 9		19	15	11	NILL	NILL
MRSA 10		24	16	16	15	NILL
MRSA 11		26	19	15	12	17
MRSA 12		18	12	10	NILL	NILL
MRSA 14		19	14	11	10	NILL
MRSA 15		20	17	15	12	10

Key: NILL-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 5: Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from Burn wound.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 20	Leaf	14	9	10	NILL	NILL
MRSA 22		15	12	10	8	7
MRSA 10		12	9	NILL	NILL	NILL
MRSA 2		NI	NI	NI	NI	NI
MRSA 7		NI	NI	NI	NI	NI
MRSA 8		NI	NI	NI	NI	NI
MRSA 5		NI	NI	NI	NI	NI
MRSA 6		NI	NI	NI	NI	NI
MRSA 1		NI	NI	NI	NI	NI
MRSA 20		21	18	15	14	NI
MRSA 22		23	21	16	10	NI
MRSA 10		18	15	13	9	9
MRSA 2		16	11	9	NI	NI
MRSA 7		22	20	17	11	NI
MRSA 8		25	19	15	12	9
MRSA 5		15	13	10	NI	NI
MRSA 6		19	18	11	11	NI
MRSA 1		20	14	9	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 6: Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from Burn wound.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 1	Leaf	14	12	NI	NI	NI
MRSA 3		11	NI	NI	NI	NI
MRSA 15		NI	NI	NI	NI	NI
MRSA 4		17	15	11	NI	NI
MRSA 7		19	16	13	NI	NI
MRSA 10		NI	NI	NI	NI	NI
MRSA 12		NI	NI	NI	NI	NI
MRSA 2		NI	NI	NI	NI	NI
MRSA 5		NI	NI	NI	NI	NI
MRSA 1		23	21	17	15	11
MRSA 3		18	15	13	13	NI
MRSA 15		18	14	NI	NI	NI
MRSA 4		22	17	15	NI	NI
MRSA 7		16	13	10	NI	NI
MRSA10		16	11	9	NI	NI
MRSA 12		24	19	17	16	11
MRSA 2		21	15	13	12	9
MRSA 5		19	15	12	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 7: Antibacterial activity of methanol crude extract of *Parkia biglobosa* against MRSA isolates from Decubitus ulcer.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 15	Leaf	14	11	NI	NI	NI
MRSA 16		NI	NI	NI	NI	NI
MRSA 18		NI	NI	NI	NI	NI
MRSA 3		NI	NI	NI	NI	NI
MRSA 5		17	15	14	11	NI
MRSA 8		19	17	11	10	10
MRSA 9		NI	NI	NI	NI	NI
MRSA 7		15	12	NI	NI	NI
MRSA 22		18	13	NI	NI	NI
MRSA 15		22	19	17	14	11
MRSA 16		24	16	12	10	NI
MRSA 18		18	18	14	11	9
MRSA 3		16	11	NI	NI	NI
MRSA 5		14	9	NI	NI	NI
MRSA 8		23	17	15	14	10
MRSA 9		25	21	19	17	11
MRSA 7		19	17	13	9	NI
MRSA 22		15	11	NI	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 8: Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Post-surgical wound.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 40	Leaf	17	16	15	NI	NI
MRSA 41		12	8	9	NI	NI
MRSA 30		15	8	NI	NI	NI
MRSA 35		NI	NI	NI	NI	NI
MRSA 1		18	11	NI	NI	NI
MRSA 3		NI	NI	NI	NI	NI
MRSA7		NI	NI	NI	NI	NI
MRSA 8		NI	NI	NI	NI	NI
MRSA 10		NI	NI	NI	NI	NI
MRSA40	Stem-bark	20	15	NI	NI	NI
MRSA 41		19	15	13	NI	NI
MRSA 30		NI	NI	NI	NI	NI
MRSA 35		18	11	NI	NI	NI
MRSA 1		15	13	NI	NI	NI
MRSA 3		NI	NI	NI	NI	NI
MRSA 7		NI	NI	NI	NI	NI
MRSA 8		20	12	NI	NI	NI
MRSA 10		NI	NI	NI	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 9: Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Burn wound.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 23	Leaf	14	12	NI	NI	NI
MRSA 25		12	9	NI	NI	NI
MRSA 2		15	12	8	NI	NI
MRSA 5		9	NI	NI	NI	NI
MRSA 7		NI	NI	NI	NI	NI
MRSA 10		NI	NI	NI	NI	NI
MRSA 6		NI	NI	NI	NI	NI
MRSA 3		14	NI	NI	NI	NI
MRSA 16		11	NI	NI	NI	NI
MRSA 23	Stem-bark	18	15	NI	NI	NI
MRSA 25		20	19	15	NI	NI
MRSA 2		15	9	NI	NI	NI
MRSA 5		NI	NI	NI	NI	NI
MRSA 7		NI	NI	NI	NI	NI
MRSA 10		NI	NI	NI	NI	NI
MRSA 6		NI	NI	NI	NI	NI
MRSA 3		NI	NI	NI	NI	NI
MRSA 16		NI	NI	NI	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 10: Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Diabetic foot ulcers.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 2	Leaf	NI	NI	NI	NI	NI
MRSA 4		NI	NI	NI	NI	NI
MRSA 8		17	14	12	NI	NI
MRSA 10		12	NI	NI	NI	NI
MRSA 6		NI	NI	NI	NI	NI
MRSA 1		NI	NI	NI	NI	NI
MRSA 14		NI	NI	NI	NI	NI
MRSA 15		15	NI	NI	NI	NI
MRSA 12		20	15	NI	NI	NI
MRSA 2	Stem-bark	NI	NI	NI	NI	NI
MRSA 4		NI	NI	NI	NI	NI
MRSA 8		12	9	NI	NI	NI
MRSA 10		NI	NI	NI	NI	NI
MRSA 6		NI	NI	NI	NI	NI
MRSA 1		17	15	15	8	NI
MRSA 14		NI	NI	NI	NI	NI
MRSA 15		NI	NI	NI	NI	NI
MRSA 12		NI	NI	NI	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

Table 11: Antibacterial activity of methanol crude extract of *Newbouldia laevis* against MRSA isolates from Decubitus ulcer.

Strain	Plant part	Extract Concentration				
		100mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	6.25 mg/ml
MRSA 13	Leaf	NI	NI	NI	NI	NI
MRSA 16		NI	NI	NI	NI	NI
MRSA 2		NI	NI	NI	NI	NI
MRSA 7		NI	NI	NI	NI	NI
MRSA 17		NI	NI	NI	NI	NI
MRSA 21		NI	NI	NI	NI	NI
MRSA 18		NI	NI	NI	NI	NI
MRSA 11		NI	NI	NI	NI	NI
MRSA 13	Stem-bark	18	13	NI	NI	NI
MRSA 16		21	17	14	11	9
MRSA 2		NI	NI	NI	NI	NI
MRSA 7		NI	NI	NI	NI	NI
MRSA 17		NI	NI	NI	NI	NI
MRSA 21		NI	NI	NI	NI	NI
MRSA 18		17	15	11	9	NI
MRSA 11		20	NI	NI	NI	NI

Key: NI-No inhibition, MRSA-Methicillin Resistant *Staphylococcus aureus*.

50-69 years and has been found elsewhere [5, 39]. This low isolation rate amongst age 50-69 years may be linked to sample size recruited in the study.

Previous antibiotic intake Assessment among wound patients showed 23.5% occurrence rate of MRSA amongst ampiclox users followed by Ampicillin users 14.5%, while least occurrence rate 7.5% was recorded against ciprofloxacin users. Ampiclox and ampicillin is the most commonly prescribed antibiotics medications to wound patients in Nigeria. MRSA strain are known to possess low affinity to β -lactam antibiotics. Therefore, exposure to sublethal dose of this β -lactam antibiotics may result in observed increase of MRSA colonization in those users. The finding in this study that MRSA is connected with key risk variables such as age, gender, antibiotic overuse, and prolonged stay in ICU has also been documented in other studies [2, 5, 33, 36, 39]. Despite considerable efforts to manage antibiotic resistance bacteria by rigorous infection control strategies, antibiotic resistant Staphylococci, particularly MRSA, have become the most common bacteria encountered in bone and wound infections in orthopaedic treatment [1, 31].

MRSA is a big concern worldwide [2], with a high significant morbidity and mortality rate in immunocompromised patients, particularly burn wound patients [4].

This study shows that MRSA exhibit resistant to macrolides (that induce erm expression) and lincosamide. This isolates demonstrate 51%-100% resistant to lincomycin and erythromycin and it's similar to another study [2, 4, 5, 7, 39]. This means that both lincomycin and erythromycin are ineffective in these patients.

Majority of our MRSA showed 100% resistant to penicillin and tetracycline and is in accordance with the pattern in Abakaliki and Jiangxi province [7, 30]. This could be due to early exposure of these isolates to this antibiotic, which may have accelerated resistance development. In our country, there is a significant degree of antibiotic abuse due to self-medication, which is frequently connected with inappropriate dosage and failure to comply with treatment, as well as the accessibility of antimicrobial agents for patients over the counter without or with a prescription.

Like other studies conducted in the Nigeria [2, 20, 23, 40] this study confirms the presence of vancomycin-resistant MRSA amongst wound patient in Abakaliki. Low vancomycin-resistant MRSA from wound 11.0% in Asmara, Eritrea [5] has been published. In contrast, several studies found that MRSA is 50-100% susceptible to vancomycin. [26, 39, 41, 42]. According to Falagas et al. [43], the sensitivity of MRSA identified in Africa to VRSA ranges between 82-100% [43]. These estimates and findings contradict an earlier report that VRSA strains are rare and there is limited indication of rising prevalence [44].

The observed heterogeneity could be explained by changes in antibiotic prescription patterns between countries. Additionally, the vanA gene complex, which encodes high degree glycopeptide resistance in enterococci, was detected in the vancomycin resistant *Staphylococcus aureus* (VRSA) isolates [2]. To date, all VRSA isolates found in the United States are methicillin resistant and carry the *mecA* gene. The vanA gene complex appears to confer alterations in the cell wall comparable to those seen in vancomycin-resistant enterococci in *S. aureus*, and *mecA* and *vanA* appear to function independently [2].

MRSA from wound in this study develops resistance to oxacillin, cephalosporins, macrolides, lincosamides, glycopeptide and Trimethoprim-Sulfamethoxazole. MRSA evolution has been linked to the acquisition of the exogenous gene (*mecA*), which is part of the staphylococcal cassette chromosome *mec* (SCCmec) (types I-VII) and is controlled by *MecI* (a repressor) and *MecR1* (a transducer) and represents the regulatory/signalling proteins of the *blaZ* system [2, 8]. The *mecA* gene encodes an extra penicillin-binding protein (PBP2a),

a peptidoglycan transpeptidase that can give resistance to all -lactam antibiotics and others antibiotic category [2, 8] as evidence in this study.

Interestingly, all MRSA our isolates showed high susceptibility to imipenem 100% while the isolates were 87.8-100% susceptible to Amikacin. In corroboration with earlier literature MRSA 73.2% and 92.6% susceptibility to imipenem and Amikacin has been reported [41, 45]. The susceptibility of this aminoglycoside and Carbapenem (regarded as last-resort antibiotic for Gram positive and negative bacterial infections) may result from their relative expensive and unavailability of this drugs for abuse. As such, imipenem and Amikacin could be considered for treatment of wound infection harboring MRSA.

Considerably, a substantial quantity of the toxin *tst* gene that mediate toxic shock syndrome in wound patients and *PVL* operon (*lukS-PV* and *lukF-PV* genes) has been reported from wound infection [33, 46]. Expression of this virulent genes may increase delay wound healing which lead to prolong hospitalization and increase duration of antibiotic administration in wound patient. The profile of multidrug antibiotic resistance reported in wound samples in this study is in line with report in the same setting [1, 5]. This implies that the isolates emerged from a high-risk source of contamination, where antibiotics are frequently administered and probably abused as a result of poor infection control and prevention policies. As such, the high Multi-drug Resistant value demonstrated among MRSA isolate may also be linked to external source of contamination beside the hospital setting in the studied patients.

Plant derived antimicrobial compounds are effectively utilized in disease management globally. Keeping the above fact in view, the antibacterial activity of *P. biglobosa* leaf and stem-bark methanol extracts show concentration dependent pattern on the MRSA isolates. *P. biglobosa* stem-bark produce inhibition zone diameter of 14mm at 25mg/ml concentration against MRSA isolates from Decubitus ulcer patients which is corroborate with the work of Obajuluwa et al. [20] who reported that twenty eight (28) MRSA isolates from bed-sore had inhibitory zone of 11-14mm at 25mg/ml concentration. Stem-bark and leaf extract exhibit high inhibitory effect of 25mm and 19mm respectively at 100mg/ml against MRSA isolates. This finding revealed that stem-bark extract had high antibacterial activity than leaf extract. Although variability in zone of inhibition diameter was observed at different concentration. The variations were comparable to prior research on *P. biglobosa* methanol leaf extract [20, 47, 48]. In accordance with the observed inhibitory activity, *Parkia biglobosa* has been found to be contain secondary metabolites e.g., tannins, flavonoids, and saponins [47], which possesses high antibacterial properties. According to Millogo-Kone et al. [15], the stem bark contains triterpenes, sterols, tannins, anthocyanins, saponosides, coumarins, reducing compounds and flavonoids, whilst the leaf contains flavonoids, coumarins, tannins, coumarins, reducing compounds and anthocyanins [15]. The higher antibacterial activity of *Parkia biglobosa* stem-bark extract found in the present research was attributed to the presence of many constituent groups that may be functioning synergistically against MRSA isolate.

From the results, most MRSA isolate were not susceptible to methanol leaf and stem bark *Newbouldia laevis* extract. It was noted that, for fewer MRSA isolate, the inhibitory effects became more pronounced at 100mg/ml and 50mg/ml as presented in the result. In contrast with this findings, previous literature has reported antibacterial activity of stem bark *Newbouldia* against *Staphylococcus aureus* from infected wound and eyes. Our result, although inconsistent with reports from previous study [10], the composition of phytochemicals in the extracts may have been changed directly by changes in the extraction process, nature, and selection of solvents utilized across investigations. This could explain the observed discrepancy. Com-

paratively, the general variability observations amongst *P. biglobosa* and *Newbouldia laevis* against MRSA in this study can be attributed to phytomolecules' affinity for solvents. The bioactive compounds found in *P. biglobosa* plant extract are thus more soluble in methanol than those found in *Newbouldia laevis*. The methanol extracts of the stems and barks were more active than the extracts of the leaves. This gap can be explained by differences in a plant's physiological features depending on its environmental microclimate, maturity and soil type. These variables are critical in the production of chemical principles and thereby influence plant pharmacological action.

Conclusion

The present study report that MRSA isolate constituted a percentage frequency of 55.5% recovered from various wound samples. In vitro failure of most antibiotic (erythromycin, lincomycin, tetracycline, Trimethoprim-Sulfamethoxazole, Vancomycin, penicillin, ceftazidime) with resistant trend of 50-100% against MRSA in this study justifies the need for more effective antibiotic therapy for wound infections with MRSA. Amikacin and imipenem were the only antimicrobial drugs that demonstrated 87.8-100% sensitivity to the test isolate, making them a better therapeutic option in single or combination regimen treatment. In comparison to *P. biglobosa* leaf and stem extract, *Newbouldia laevis* leaf and stem bark methanol extract had no good inhibitory effect at 25mg/ml, 12.5mg/ml, and 6.25 mg/ml concentrations against MRSA isolate. As a result, the findings of this study have verified folklore medicine's assertions about the use of *P. biglobosa* for the treatment of infections such as wounds. The ample presence of phytochemicals substance in the methanol extracts *P. biglobosa* underscore their antibacterial activity against MRSA isolates in this study. The *Newbouldia laevis* plant's limited or low antibacterial activity in vitro need further testing with different solvents. More research with a wide range of target strains is required to determine

Reference

1. Udobi, C. E., Obajuluwa, A. F and Onaolapo, J. A (2013). Prevalence and Antibiotic Resistance Pattern of Methicillin-Resistant *Staphylococcus aureus* from an Orthopaedic Hospital in Nigeria, Hindawi Publishing Corporation BioMedical Research International, 12(4): 467-860.
2. Peter IU, Okolo IO, Uzoeto HO, Edemekong CI, Thompson MD, Chukwu EB, Mohammed, ID, Ubom, IJ, Joseph, OV, Nwuzo, AC, Akpu PO, Iroha, IR (2022a). Identification and antibiotic resistance profile of biofilm-forming Methicillin Resistant *Staphylococcus aureus* (MRSA) causing infection among orthopedic wound patients. Asian J Res Med Pharm Sci. 2022a;11(4):45-55.
3. Peter IU, Ngwu JN, Edemekong CI, Ugwueme IV., Uzoeto HO., Joseph OV, Mohammed ID., Mbong EO, Nomeh OL, Ikusika BA., Ubom IJ., Inyogu JC., Ntekpe ME., Obodoechi IF., NseAbasi, PL., Ogbonna IP., Didiugwu CM, Akpu PO., Alagba EE., Ogbu RC, Iroha, IR. First report prevalence of Livestock Acquired Methicillin Resistant *Staphylococcus aureus* (LA-MRSA) strain in South Eastern, Nigeria . IOSR J Nurs Health Sci. 2022b;11(1):50-56.
4. Ghaznavi-Rad, E and Ekrami, A (2018).Molecular Characterization of Methicillin-Resistant *Staphylococcus aureus* Isolates, Isolated from a Burn Hospital, in Southwest Iran in 2006 and 2014, Hindawi International Journal of Microbiology, 5(23):123-200.
5. Garoy, E. Y., Gebreab, Y. B., Achila, O. O., Tekeste, D. G., Robel, R. K., Kiflay, G. R and Tesfu, T. (2018). Methicillin-Resistant *Staphylococcus aureus* (MRSA): Prevalence and Antimicrobial Sensitivity Pattern among Patients—A Multicenter Study in Asmara, Eritrea, Canadian Journal of Infectious Diseases and Medical Microbiology,9: 832-1834.
6. Holmes, A., Ganner, M., McGuane, S., Pitt, T. L., Cookson, B. D and Kearns, A. M (2005). *Staphylococcus aureus* isolates carrying Panton-Valentine leucocidin genes in England and Wales: frequency, characterization, and association with clinical dis-
- ease, Journal of Clinical Microbiology, 43(5):2384–90.
7. Chen., K., Lin, S., Li, P., Song, Q., Luo, D., Liu, T., Zeng, Land Zhang, W (2018). Characterization of *Staphylococcus aureus* isolated from patients with burns in a regional burn center, Southeastern China, Biomedical Complement of Infectious Diseases,18:51-60.
8. Peter IU, Edemekong C, Balogun OM, Ikusika BA, Ngwu JN, Bassey NU, Mohammed ID, et al. Phytochemical Screening and Antibacterial Activity of Aqueous Extract of *Costus afer* Plant on Selected Multidrug-Resistant Bacteria. IOSR J Pharmacy and Biol Sciences. 2022c; 17(1):39-42.
9. Ude IU, Moses IB, Okoronkwo C, Ovia K, Okafor C, Chukwunwejim CR, OkataNwali OD, Iroha CS, Akuma S, Peter IU, Uzoeto HO, et al. Phytochemical properties and antimicrobial activity of *Buchholzia coriacea* and *Psychotria microphylla* leaf extracts on bacterial pathogens isolated from aquatic environments in Nigeria. J Medicinal Plants Res, 2021; 15(6):232-240.
10. Akerele, J. O., Ayinde, B. A and Ngiagah, J (2011), Phytochemical and Antibacterial Evaluations of the Stem Bark of *Newbouldia laevis* against Isolates from Infected Wounds and Eyes, Tropical Journal of Pharmaceutical Research, 10 (2): 211-217.
11. Nwode V. Detection of Methicillin Resistant *Staphylococcus aureus* and associated risk factors among wound patient in a Tertiary Hospital in Abakaliki, (M. Sc Thesis at Ebonyi state University, Abakaliki; 2021.
12. Germann, K., Kaloga, M., Ferreira, D., Marais, J. P and Kolodziej, H (2006). Newbouldioside A-C Phenylethanoid Glycosides from the Stem-bark of *Newbouldia laevis*. Journal of Phytochemistry, 67(8): 805–811.
13. Usman, H and Osuji, J. C (2007). Phytochemical and In vitro Antimicrobial Assay of the Leaf Extract of *Newbouldia laevis*. African Journal of Traditional, Complementary and Alternative Medicine, 4(4): 476-480.

the abundant pharmacological value of the *Newbouldia laevis* and *P. biglobosa* plant on a large scale.

Abbreviations

MRSA	Methicillin Resistant <i>Staphylococcus aureus</i>
NI	No Inhibition
AST	Antibiotic Susceptibility Testing

Funding

There is no source of funding to declare

Author's contribution

This work was carried out in collaboration among all authors. Authors VN, KA wrote the protocol. Authors IUP and IRI wrote the first draft of the manuscript. Authors LU, IO and HO managed the characterization and analyses of the study. All authors read and approved the final manuscript.

Acknowledgements

The authors sincerely acknowledge the patients and Medical Staff of Alex Ekwueme Federal University, Abakaliki for their support.

Competing Interests

Authors have declared that no competing interests exist.

Data availability Statement

Data supporting these findings are available within the article or upon request

Institutional review board statement

The ethical committee of Alex Ekwueme Federal University Teaching Hospital in Abakaliki, Ebonyi State, authorized this study conveyed with approval number: 26/11/2020. This was based on a thorough knowledge of the scientific literatures, satisfactory laboratory protocols, and other relevant sources of information guiding this area of research. Every fundamental study was done in line with the World Medical Association (WMA) declaration of Helsinki on the principles for medical research involving human subjects, and identifiable human material or data.

14. Jauro, S., Abubakar, M. B., Geidam, Y. A., Zanna, M. Y., Kwoji, I. D., Isa-Gulani, A and Ibrahim, I (2018). Phytochemical and Antimicrobial Profile Analysis of *Parkia biglobosa* against Methicillin-Resistant *Staphylococcus aureus*. *Journal of Advanced Veterinary and Animal Research*, 5(2):173-181.
15. Millogo-Kone, H., Guissou, I. P., Nacoulma, O and Traore, A. S (2007). Antimicrobial effects of the stem Bark extracts of *Parkia biglobosa* on *Shigellae*. *African Journal of Traditional, Complementary and Alternative Medicine*, 4:392-396.
16. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing; twenty-eighth edition (M100). Wayne, PA: Clinical and Laboratory Standards Institute; 2019.
17. Osuntokun, O. T., Jemilaiye, T. A and Akinrodoye, A. R (2018). Comparative Study between the Effect of *Parkia Biglobosa* (JACQ) benth and Conventional Antibiotics against Multiple Antibiotic Resistant Uropathogenic Bacteria (MARUB). *Journal of Bioequivalence and Bioavailability*, 5(4):200-212.
18. Oyama, M. O., Egbebi, A. O and Akharaiyi, F. C (2018). Phytochemical Analysis and Antibacterial Activities of Some plant Extracts on *Staphylococcus aureus* isolates from Patients Receiving Hospital Treatments in Ekiti State, Nigeria. *Journal of Herbal medicine and Pharmacology*, 9(1):14-20.
19. Olowe, O. A., Eniola, K. I. T., Olowe, R. A and Olayemi, A. B (2007). Antimicrobial Susceptibility and Beta-lactamase detection of MRSA in Osogbo, SW Nigeria. *Nature and Science*, 5(3):44-48.
20. Obajuluwa, A. F., Onaolapo, J. A., Oyi, A. R and Olayinka, B. O (2013). Susceptibility Profile of Methicillin-Resistant *Staphylococcus aureus* (MRSA) Isolates to Antibiotics and Methanolic Extracts of *Parkia biglobosa* (Jacq) Benth. *British Journal of Pharmaceutical Research*, 3(4): 587-596.
21. Ikeh, E. C (2003). Methicillin-resistant *Staphylococcus aureus* at Jos University Teaching Hospital. *African Journal of Clinical and Experimental Microbiology*, 4(1):52-62.
22. Nwankwo, E. O and Nasiru, M. S (2011). Antibiotic Sensitivity Patterns of *Staphylococcus aureus* from Clinical Isolates in a Tertiary Health Institution in Kano, Northwestern Nigeria. *Pan African Medical Journal*, 8(4):123-131.
23. Taiwo, S. S., Onile, B. A and Akanbi, A. A (2004). Methicillin resistant *Staphylococcus aureus* (MRSA) Isolates in Ilorin, Nigeria. *African Journal of Clinical and Experimental Microbiology*, 5(2):189-197.
24. Mawalla, B., Mshana, S. E., Chalya, P. L., Imirzalioglu, C and Mahalu, W (2011). Predictors of Surgical Site Infections Among Patients Undergoing Major Surgery at Bugando Medical Centre in Northwestern Tanzania. *BioMedical Complement of Surgical Infection*, 11: 21-25.
25. Anguzu, J. R and Olila, D (2007). Drug Sensitivity Patterns of Bacterial Isolates from Septic Post-operative Wounds in a Regional Referral Hospital in Uganda. *African Journal of Health Science*, 7:148-154.
26. Ghanem, S., Bahashwan, S. A., El Shafey, H. M., Fayed, A. A., Alhhazmi, A and Manzoor, N (2018). Antimicrobial resistance pattern of MRSA strains isolated from patients of a hospital in Madinah, Kingdom of Saudi Arabia, *African Journal of Microbiology Research*, 12(47):1044-1049.
27. Rebiah, S. A., Abdelouahid, D. E., Rahmoun, M., Abdelali, S and Azzaoui, H (2011). Emergence of vancomycin-resistant *Staphylococcus aureus* Identified in the Tlemcen University Hospital (North-West Algeria). *Médecine et Maladies Infectieuses*, 41(12):646-651.
28. Guzman-Blanco, M., Carlos, M and Raul, I (2009). Review Epidemiology of Methicillin-resistant *Staphylococcus aureus* (MRSA) in Latin America. *International Journal of Antimicrobial Agents*, 34(4): 304-308.
29. Jimenez, J. N., Ocampo, A. M and Vanegas, J. M (2012). CC8 MRSA Strains Harboring SCCmec type IV are Predominant in Colombian Hospitals. *Public Library of Science One*, 7(6):38-576.
30. Ariom, T. O., Iroha, I. R., Moses, I. B., Iroha, C. S., Ude, U. I and Kalu, A. C (2019). Detection and phenotypic characterization of methicillin-resistant *Staphylococcus aureus* from clinical and community samples in Abakaliki, Ebonyi State, Nigeria, *African Journal of Health Science*, 19(2):2026-2035.
31. Peter IU, Emelda NC, Chukwu EB, Ngwu JN, Uzoeto HO, Moneth EC, Stella AO, Edemekong CI, Uzoamaka EP, Nwuzo AC, Iroha IR. Molecular detection of bone sialoprotein-binding protein (bbp) genes among clinical isolates of methicillin resistant *Staphylococcus aureus* from hospitalized orthopedic wound patients. *Asian J Orthopaedic Res*. 2022d;8(3):1-9.
32. Deyno, S., Fekadu, S and Astatkie, A (2017). Resistance of *Staphylococcus aureus* to Antimicrobial Agents in Ethiopia: a Metaanalysis. *Antimicrobial Resistance and Infection Control*, 6(1):85-17.
33. Kolawole, D. O., Adeyanju, A., Schaumburg, F., Akinyoola, A. L and Lawal, O. O (2013) Characterization of Colonizing *Staphylococcus aureus* Isolated from Surgical Wards' Patients in a Nigerian University Hospital. *Public Library of Science One*, 8(7):68-721.
34. Shirah, B. H., Zafar, S. H., Alferaidi, O. A and Sabir, A. M. M (2017). Mass Gathering Medicine (Hajj pilgrimage in Saudi Arabia): The Clinical Pattern of Pneumonia Amongst Pilgrims During Hajj. *Journal of Infection and Public Health*, 10(3):277-286.
35. Ali, S. M., Bayoumi, E. A. M and Alshazly, T. A (2013). Molecular Identification of Methicillin-resistant *Staphylococcus aureus* and Evaluation of Panton-valentine leukocidin Gene as a Sole Marker for CA-MRSA. *Journal of American Science*, 9(8):108-115.
36. Haseeb, A., Faidah, H. S., Bakhsh, A. R., Malki, W. H., Elrggal, M. E., Saleem, F., Rahman, S. U., Khan, T. M and Hassali, M. A (2016). Antimicrobial Resistance Amongst Pilgrims: A Retrospective Study from Two Hospitals in Makkah, Saudi Arabia. *International Journal of Infectious Diseases*, 47:92-94.
37. Magliano, E., Grazioli, V., Deflorio, L., Leuci, A. I., Mattina, R and Romano, P (2012). Gender and Age-dependent Etiology of community-acquired urinary tract infections. *Scientific World Journal*, 16(12):123-456.
38. Neuman, H., Debelius, J. W., Knight, R and Koren, O (2015). Microbial Endocrinology: The Interplay between the Microbiota and The Endocrine System. *Federation of European Medical Societies of Microbiology Reviews*, 39(4):509-521.
39. Seni, J., Bwanga, F., Najjuka, C. F., Makobore, P and Okee, M (2013). Molecular Characterization of *Staphylococcus aureus* from Patients with Surgical Site Infections at Mulago Hospital in Kampala, Uganda. *Public Library of Science One*, 8(6):66-153.
40. Onolitola, O. S., Olayinka, B. O., Salawu, M. J and Yakubu, S. E (2007). Nasal carriage of methicillin resistant *Staphylococcus aureus* with reduced vancomycin susceptibility (MRSA-RVS) by healthy adults in Zaria, Nigeria *Journal of Tropical Microbiology and Biotechnology*, 3:19-22.
41. Abdulla, N and Iregbu, K. C (2019). Methicillin-Resistant *Staphylococcus aureus* in a Central Nigeria Tertiary Hospital. *Annal of Tropical Pathology*, 9:6-10.
42. Omuse, G., Kariuki, S and Revathi, G (2012). Unexpected Absence of Methicillin-resistant *Staphylococcus aureus* Nasal Carriage by Healthcare Workers in a Tertiary Hospital in Kenya. *Journal of Hospital Infection*, 80(1):71-73.
43. Falagas, M. E., Karageorgopoulos, D. E., Leptidis, J and Korbila, I. P (2013) MRSA in Africa: Filling the Global Map of Antimicrobial Resistance. *Public Library of Science One*, 8(7):680-24.
44. Kong, E. F., Johnson, J. K and Jabra-Rizk, M. A (2016). Community Associated Methicillin-resistant *Staphylococcus aureus*: An Enemy Amidst Us. *Public Library of Science Pathogens*, 12(10):100-5837.
45. Rajaduraipandi, K., Mani, K. R., Panneerselvam, K., Mani, M.,

- Bhaskar, M and Manikandan, P (2006). Prevalence and Antimicrobial Susceptibility Pattern of Methicillin Resistant Staphylococcus aureus: A Multicentre Study. Indian Journal of Medical Microbiology, 24:34-38.
46. Shittu, A. O., Okon, K., Adesida, S., Oyedara, O and Witte, W (2011) Antibiotic Resistance and Molecular Epidemiology of Staphylococcus aureus in Nigeria. BioMedical Complement of Microbiology, 11: 92-100.
47. Ajaiyeoba, E. O (2002). Phytochemical and Antibacterial Properties of Parkia biglobosa and Parkia bicolor leaf extracts. African Journal of Biomedical Research, 5(3):125–129.
48. Udobi, C. E., Onaolapo, J A and Agunu, A (2010). Antibacterial Potentials of the Methanolic Extract and Aqueous Fraction of the Stem Bark of the African Locust Bean (Parkia Biglobosa). European Journal of Scientific Research, 43(4):590–597.